

## L form Case Definitions 2024

Sl. No.	Disease Condition	Case Definition
1.	<b>Influenza</b>	<p>A presumptive case of ILI or SARI with</p> <ul style="list-style-type: none"> <li>• Conventional Polymerase Chain Reaction (PCR) or real-time reverse transcription PCR (RT-PCR)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Any validated nucleic acid-based test (Annexure 1)</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, Technical Guidelines on H1N1, 2019</i></p> </div>
2.	<b>Cholera</b>	<p>A presumptive case of acute diarrheal disease with</p> <ul style="list-style-type: none"> <li>• Culture (and identification of <i>Vibrio cholera</i>)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Polymerase Chain Reaction (PCR) test</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: WHO–recommended standards for Surveillance of selected Vaccine-Preventable Diseases, 2018</i></p> </div>
3.	<b>Shigellosis</b>	<p>An acute diarrhea / dysentery case with</p> <ul style="list-style-type: none"> <li>• Isolation of <i>Shigella</i> species from stool sample (Culture)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Nucleic acid-based tests</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Public Health Laboratory Network case definitions, May 2000</i></p> </div>
4.	<b>Typhoid</b>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Positive culture (from any clinical specimen)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Molecular methods of <i>S. typhi</i>/ <i>S. paratyphi</i></li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: WHO–recommended standards for Surveillance of selected Vaccine-Preventable Diseases, 2018</i></p> </div>

5.	<b>Hepatitis A</b>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• IgM antibodies to Hepatitis A (anti HAV IgM) in serum/plasma</li> </ul> <p><i>Note: The sample has to be tested as per testing algorithm for jaundiced patient according to guidelines of National Viral Hepatitis Control Programme.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: National Viral Hepatitis Control Programme shared on 10.06.2019</i></p> </div>
6.	<b>Hepatitis E</b>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• IgM antibody to Hepatitis E virus (anti HEV IgM) in serum/plasma</li> </ul> <p><i>Note: The sample has to be tested as per testing algorithm for jaundiced patient according to guidelines of National Viral Hepatitis Control Programme</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: National Viral Hepatitis Control Programme shared on 10.06.2019</i></p> </div>
7.	<b>Meningitis (Meningococcal disease)</b>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Antigen detection by Latex Agglutination Test in CSF</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Isolation of N. meningitides from blood or CSF</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Detection of N. meningitides-specific nucleic acid in a specimen (e.g., blood or CSF), using a validated polymerase chain reaction (PCR) assay</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert November,2009</i></p> </div>

<p><b>8.</b></p>	<p><b>Diphtheria</b></p>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Isolation of <i>Corynebacterium diphtheriae</i> from a clinical specimen by culture</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Detection by PCR from a clinical specimen</li> </ul> <p><i>Note: Throat swab/pieces of membrane collection within 4 weeks of onset of sore throat.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Immunization Division shared on 13.10.2023</i></p> </div>
<p><b>9.</b></p>	<p><b>Pertussis</b></p>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Isolation of Bordetella pertussis from a clinical specimen by culture</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Detection by PCR from a clinical specimen</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Single serum positive for IgG antibody</li> </ul> <p><i>Note: Nasopharyngeal swab collection within 4 weeks of onset of cough and serology sample collection within 12 weeks of onset of cough.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Immunization Division shared on 13.10.2023</i></p> </div>

<b>10.</b>	<b>Measles</b>	<p>A presumptive case with</p> <ul style="list-style-type: none"><li>• Detection of anti-measles IgM antibody by enzyme immunoassay (EIA)</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• Measles virus detection through PCR from throat swab or urine or nasopharyngeal swab</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• Isolation of measles virus</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• Direct epidemiologic linkages to a case confirmed by one of the above methods</li></ul> <p><i>Note: Virology sample collection within 7 days and serology sample collection within 28 days of onset of rash.</i></p> <div data-bbox="531 630 1528 683" style="border: 1px solid black; padding: 2px;"><p><i>Source: Immunization Division shared on 13.10.2023</i></p></div>
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<p><b>11.</b></p>	<p><b>Rubella</b></p>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Detection of anti-rubella IgM antibody by enzyme immunoassay (EIA)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Rubella virus detection through PCR from throat swab or urine or nasopharyngeal swab</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Isolation of rubella virus</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Direct epidemiologic linkages to a case confirmed by one of the above methods</li> </ul> <p><i>Note: Virology sample collection within 7 days and serology sample collection within 28 days of onset of rash.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Immunization Division shared on 13.10.2023</i></p> </div>
<p><b>12.</b></p>	<p><b>Polio</b></p>	<p>An AFP case is ‘confirmed’ as polio only by the</p> <ul style="list-style-type: none"> <li>• Isolation of wild poliovirus from any stool specimen in the WHO accredited laboratory</li> </ul> <p><i>Note: Stool sample collection within 14 days of paralysis onset, and no later than 60 days following paralysis onset.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Immunization Division shared on 13.10.2023</i></p> </div>

<b>13.</b>	<b>Mumps</b>	<p>A presumptive case</p> <ul style="list-style-type: none"><li>• Positive by Reverse Transcription-Polymerase Chain Reaction (RT-PCR) from an appropriate clinical specimen (buccal/oral swab, throat swab, urine, and cerebrospinal fluid)</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• Seroconversion from IgG negative to IgG positive as determined by any standard serological assay in the absence of mumps immunization in the preceding six weeks</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• In unvaccinated individuals, significant (<math>\geq</math> fourfold) rise in serum mumps IgG titre as determined by any standard serological assay</li></ul> <p><b>OR</b></p> <ul style="list-style-type: none"><li>• Isolation of mumps virus by culture from an appropriate clinical specimen (buccal/oral swab, throat swab, urine, and cerebrospinal fluid)</li></ul> <p><i>Source: WHO–recommended standards for Surveillance of selected Vaccine-Preventable Diseases,2018 (Modified on 28.05.2019, NCDC)</i></p>
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<p><b>14.</b></p>	<p><b>Chicken pox</b></p>	<p>A presumptive case with</p> <ul style="list-style-type: none"> <li>• Detection of VZV DNA by Polymerase Chain Reaction (PCR)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Direct antigen detection of VZV from an appropriate clinical specimen {e.g. direct fluorescent antibody (DFA) testing}</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Virus isolation</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Seroconversion or a significant rise (fourfold or greater) in varicella-zoster IgG titer between acute and convalescent sera by any validated serologic assay</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: WHO–recommended standards for Surveillance of selected Vaccine-Preventable Diseases,2018 (Modified on 28.05.2019, NCDC)</i></p> </div>
<p><b>15.</b></p>	<p><b>Malaria</b></p>	<p>A confirmed malaria case (or infection) is one in which the parasite has been detected by a diagnostic test, i.e. microscopy,rapid diagnostic test, or molecular diagnostic test.</p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCVBDC shared on 22.09.2023</i></p> </div>

<p><b>16.</b></p>	<p><b>Dengue</b></p>	<p>A case compatible with the clinical description (see below) of Dengue Fever with at least one of the following:</p> <ul style="list-style-type: none"> <li>• Isolation of dengue virus (Virus culture+VE) from serum, plasma, leucocytes</li> <li>• Demonstration of IgM antibody titre by ELISA positive in single serum sample</li> <li>• Demonstration of dengue virus antigen in serum sample by NS1-ELISA</li> <li>• IgG seroconversion in paired sera after 2 weeks of fourfold increase of IgG titre</li> <li>• Detection of viral nucleic acid by polymerase chain reaction (PCR)</li> </ul> <p>Clinical Description of Dengue: The clinical description of Dengue Fever includes an acute febrile illness of 2-7 days duration with two or more of the following manifestations:</p> <ul style="list-style-type: none"> <li>• Headache</li> <li>• Retro-orbital pain</li> <li>• Myalgia</li> <li>• Arthralgia</li> <li>• Rash</li> <li>• Haemorrhagic Manifestations</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCVBDC shared on 22.09.2023</i></p> </div>
<p><b>17.</b></p>	<p><b>Chikungunya</b></p>	<p>A patient meeting both the clinical and laboratory criteria:</p> <p><b>Clinical criteria:</b> Acute onset of fever and severe arthralgia/arthritis with or without skin rash and residing or having left an epidemic area 15 days prior to onset of symptoms</p> <p><b>Laboratory criteria:</b> At least one of the following tests done in the acute phase of illness</p> <p><b>Direct evidence:</b> Virus isolation / Presence of viral RNA by RT-PCR</p> <p><b>Indirect evidence:</b></p> <ul style="list-style-type: none"> <li>• Presence of virus specific IgM antibodies in single serum sample collected in acute or convalescent stage.</li> <li>• Four-fold increase in IgG values in samples collected at least three weeks apart.</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCVBDC shared on 22.09.2023</i></p> </div>



18.	<b>Japanese Encephalitis</b>	<p>Every AES is a Suspect case of JE and it has to be laboratory confirmed <b>with any one</b> of the following markers:</p> <ul style="list-style-type: none"> <li>• Presence of IgM antibody in serum and/ or CSF to JE Virus</li> <li>• Fourfold difference in IgM antibody titre in paired sera</li> <li>• Virus isolation from brain tissue</li> <li>• Antigen detection by immunofluorescence</li> </ul> <p>Nucleic acid detection by PCR</p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCVBDC shared on 22.09.2023</i></p> </div>
19.	<b>West Nile Virus</b>	<p>An Acute Encephalitic Syndrome (AES) case with:</p> <ul style="list-style-type: none"> <li>• Viral detection by Reverse Transcription Polymerase Chain Reaction (RT-PCR) assay,</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• IgM antibody capture enzyme-linked immunosorbent assay (ELISA); PRNT is recommended to rule out cross reactions and confirmation.</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Virus isolation by cell culture.</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Updated by Zoonosis Division NCDC on 02.07.2019</i></p> </div>
20.	<b>Kala-azar</b>	<p>A presumptive case is confirmed by:</p> <ul style="list-style-type: none"> <li>• Rapid diagnostic test (RDT)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Polymerase Chain Reaction (PCR)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Biopsy</li> </ul> <p><i>#In cases with past history of kala-azar or with high suspicion and with negative RDT results, confirmation can be done by examination of bone marrow/spleen aspirate for Leishmania donovani (LD) bodies at an appropriate level equipped with such skills and facilities.</i></p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCVBDC shared on 26.09.2023</i></p> </div>

<p><b>21.</b></p>	<p><b>Human Rabies</b></p>	<p>A suspected or probable case that is laboratory-confirmed*</p> <p>*Laboratory confirmation is done by one or more of the following:</p> <ul style="list-style-type: none"> <li>• Detection of viral antigens in clinical samples (e.g., brain tissue, nuchal skin biopsy, saliva)</li> <li>• Detection of viral nucleic acids in clinical samples (e.g., brain tissue, CSF, nuchal skin biopsy, saliva, urine)</li> <li>• Detection of virus-specific antibodies in the cerebrospinal fluid (CSF)</li> <li>• Detection of virus-specific antibodies in the serum of an unvaccinated person</li> <li>• Demonstration of a four-fold or higher rise in virus-specific antibody titres in paired sera of previously vaccinated individuals</li> <li>• Isolation of rabies virus from a clinical sample (e.g. brain tissue, saliva) in cell culture or laboratory animal</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: Centre for One Health shared on 15.12.2023</i></p> </div>
<p><b>22.</b></p>	<p><b>Plague</b></p>	<p>A presumptive case with an isolate from a clinical sample identified as <i>Y. pestis</i> and two of the four following tests must be positive</p> <ul style="list-style-type: none"> <li>• <i>Y. pestis</i> biochemical profile</li> <li>• Bacteriophage lysis of culture</li> <li>• F1 Antigen detection</li> <li>• PCR (pla gene, F1 gene)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• A fourfold difference in anti F1 antibody titer in paired serum samples</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Direct validated PCR on clinical specimen</li> </ul>

<p><b>23.</b></p>	<p><b>Leptospirosis</b></p>	<p>A case compatible with the clinical description of leptospirosis with at least one of the following:</p> <ol style="list-style-type: none"> <li>1. High titre of IgM antibodies in ELISA (evaluated with locally determined cut off) for single clinical sample*.</li> <li>2. Four-fold or greater rise or persistent titre (in case of antibiotic given) in the MAT (total antibodies) between acute and convalescent-phase serum specimens run parallel.</li> <li>3. Seroconversion on ELISA in paired serology (demonstrating conversion of IgM to IgG antibodies).</li> </ol> <p><i>*A single serum sample showing high titres of IgM antibodies may indicate acute infection These 1-3 tests are the preferred tests as ELISA are widely acceptable</i></p> <p>Other: Isolation and Validated PCR can be done in patients who have not received antibiotic and in early stage of diseases (preferably less than 7 days)</p> <div style="border: 1px solid black; padding: 5px; width: fit-content; margin-left: auto; margin-right: auto;"> <p><i>Source: CAZD shared on 10.10.2023</i></p> </div>
<p><b>24.</b></p>	<p><b>Scrub Typhus</b></p>	<p>A case compatible with the clinical description of scrub typhus with at least one of the following:</p> <ol style="list-style-type: none"> <li>1. High titre of IgM antibodies in ELISA (evaluated with locally determined cut off) for single clinical sample*.</li> <li>2. A four-fold rise in the Weil-Felix test (total antibodies) between acute and convalescent-phase serum specimens run in parallel.</li> <li>3. Seroconversion on ELISA/ IFAT (demonstrating the conversion of IgM to IgG antibodies).</li> </ol> <p><i>*A single serum sample showing high titres of IgM antibodies may indicate acute infection. These 1-3 tests are the preferred tests as ELISA are widely acceptable</i></p> <p>Other: Isolation and Validated PCR can be done in patients who have not received antibiotic and in early stage of diseases (preferably less than 7 days)</p> <div style="border: 1px solid black; padding: 5px; width: fit-content; margin-left: auto; margin-right: auto;"> <p><i>Source: CAZD shared on 10.10.2023</i></p> </div>

25.	<b>Brucellosis</b>	<p>A case compatible with the clinical description of brucellosis with at least one of the following:</p> <ol style="list-style-type: none"> <li>1. High titre of IgM antibodies in ELISA (evaluated with locally determined cutoff) for single clinical sample*.</li> <li>2. A four-fold rise in the SAT (total antibodies) between acute and convalescent-phase serum specimens run parallel.</li> <li>3. Seroconversion on ELISA in paired serology (demonstrating the conversion of IgM to IgG antibodies).</li> </ol> <p><i>*A single serum sample showing high titres of IgM antibodies may indicate acute infection These 1-3 tests are the preferred tests as ELISA are widely acceptable</i></p> <p>Other: Isolation and Validated PCR can be done in patients who have not received antibiotic and in early stage of diseases (preferably less than 7 days)</p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: CAZD shared on 10.10.2023</i></p> </div>
26.	<b>Anthrax</b>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• Isolation and identification of <i>B. anthracis</i> from relevant samples and identified by colony morphology, microscopy and biochemical tests</li> <li>• Gamma phage lysis OR Validated PCR (Toxin and capsule genes) may be used for final confirmation (Validated PCR on direct clinical sample is also acceptable)</li> </ul>
27.	<b>CCHF (Crimean Congo Hemorrhagic Fever)</b>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• Detection of CCHF virus genome by validated RT - PCR in a clinical specimen AND/ OR sequencing</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Detection by ELISA or IFA of specific IgM antibodies against CCHF virus</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• A 4-fold increase in specific IgM antibodies against CCHF virus in two specimens collected in the acute and convalescence phases</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• CCHF virus isolation</li> </ul>

28.	<b>KFD (Kyasanur Forest Disease)</b>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• Detection of KFDV-specific viral RNA by reverse transcription polymerase chain reaction (RT-PCR) or real time RT-PCR from blood or tissues</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Positive for immunoglobulin M (IgM) enzyme-linked immunosorbent assay (ELISA) for KFD</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Isolation of KFDV in cell culture or in a mouse model from blood or tissues</li> </ul>
29.	<b>Nipah Virus Disease</b>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• Nipah virus RNA identified by Polymerase Chain Reaction (PCR) from respiratory secretions, urine, or cerebrospinal fluid.</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Isolation of Nipah virus from respiratory secretions, urine, or cerebrospinal fluid</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert September, 2023</i></p> </div>
30.	<b>Zika Virus Disease</b>	<p>A presumptive case with positive result for the specific detection of ZIKV by Reverse Transcription Polymerase Chain Reaction (RT-PCR)</p> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert March, 2016</i></p> </div>
31.	<b>Monkeypox</b>	<p>A presumptive case is confirmed for monkeypox virus by</p> <ul style="list-style-type: none"> <li>• Polymerase Chain Reaction (PCR)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Sequencing</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert July, 2022</i></p> </div>

32.	<b>Ebola</b>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• IgM antibody positive by enzyme-linked immunosorbent assay (ELISA)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Polymerase Chain Reaction (PCR)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Virus isolation</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert December, 2022</i></p> </div>
33.	<b>Yellow Fever</b>	<p>A presumptive case, in the absence of recent yellow fever vaccination,</p> <ul style="list-style-type: none"> <li>• Yellow-fever- specific IgM is found in the serum,</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• A fourfold or greater rise in IgG levels is found in PAIRED acute and convalescent sera,</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Yellow fever virus is isolated in cell culture or laboratory animals, or in case of positive post-mortem liver histopathology,</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Yellow fever antigens are detected in tissues by immunohistochemistry</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Yellow fever virus genomic sequences are detected in blood or organs by molecular diagnostic techniques such as Reverse Transcription Polymerase Chain Reaction (RT- PCR)</li> </ul>

<p><b>34.</b></p>	<p><b>MERS CoV</b></p>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• The presence of viral nucleic acid can be confirmed by either positive results for nucleic acid amplification assays such as: <ul style="list-style-type: none"> <li>❖ reverse transcription polymerase chain reaction (RT-PCR), for at least two specific genomic targets</li> </ul> </li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>❖ A single positive target with sequencing of a second target.</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Demonstration of sero-conversion in 2 samples ideally taken at least 14 days apart, by a screening test (ELISA) and confirmation by a microneutralization assay</li> </ul>
<p><b>35.</b></p>	<p><b>Marburg Virus Disease</b></p>	<p>A presumptive case with:</p> <ul style="list-style-type: none"> <li>• IgM antibody positive by enzyme-linked immunosorbent assay (ELISA)</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Virus isolation</li> </ul> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> <p><i>Source: NCDC, CD Alert July, 2023</i></p> </div>

## Validated Human Influenza Testing Kits (Updated on 11.04.2024\*)

<b>Kits evaluated at NIV and passed ICMR criteria</b>				
<b>Sr No</b>	<b>Manufacturer</b>	<b>Kit name</b>	<b>Virus detected</b>	<b>Remark</b>
1	Siemens HealthCare Pvt Ltd	IMDX Flu Panel (InfA/InfB/H1N1/H3N2)	Influenza A and its subtype H1N1, H3N2	
2	ALTONA DIAGNOSTICS Pvt Ltd	FlexStar® SARS-CoV-2 Type & FLU RT-PCR	SARS-CoV-2, Influenza	Kit does not differentiate between Influenza A and Influenza B.
3	Invitrogen Bioservices India Pvt. Ltd	TaqPath Covid-19, FluA, FluB Combo Kit	SARS-CoV-2, Influenza A, Influenza B	
4	M/s Kriya Medical Technologies Pvt. Ltd	KRIVIDA TRIVUS RESPI PANEL	SARS-CoV-2, Influenza A or Influenza B, RSV A or B	Kit does not differentiate between Influenza A and Influenza B, similarly RSV A & RSV B
5	HiMedia Laboratories Pvt. Ltd	Hi-PCR Influenza Multiplex Probe PCR Kit	Influenza A and its subtype H1N1, Influenza B	Kit does not detect H3N2 subtype
6	HiMedia Laboratories Pvt. Ltd	Hi-PCR COVID FLU RSV Multiplex Probe PCR Kit	Influenza A and its subtype H1N1, H3N2, Influenza B, SARS-CoV-2, RSV	Kit does not differentiate between RSV A & RSV B
7	Mylab Discovery Solutions Pvt. Ltd	Patho Detect Human Influenza Detection Kit	Influenza A and its subtype H1N1, H3N2, Influenza B	
8	<b>NIV Single tube assay for detection of SARS Cov 2</b>	<b>Technology been transferred to Mylab discovery solutions</b>		
9	<b>NIV Kits ( Multiplex single tube assay for Influenza A,B and SARS CoV 2</b>	<b>Technology transfer is in process with three companies Huwel Life sciences Meril Diagnostics Mylab solutions</b>		
<b>*Note: The list of validated Human Influenza testing kits will be updated from time to time</b>				



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